

Amendments to the Specification:

Please replace the paragraph on page 1 beginning at line 6 with the following rewritten paragraph:

This invention relates to an inducible gene expression system, particularly but not exclusively involving eukaryotes, such as plants, for example.

Please replace the paragraph on page 11 beginning at line 4 with the following rewritten paragraph:

Figure 27 shows a schematic diagram of the plasmid pOH006 as used in the present invention. The plasmid is a double construct in pBINplus, containing the chimaeric promoter (SEQ ID NO: 19), a GUS intron (Vancanneyt *et al.*, 1990) and a nos terminator, and also containing the translational fusion between the ohpR coding sequence (nucleotide 295 to nucleotide 1035 of SEQ ID NO: 19) and part of the C1 cDNA (from the NarI at nucleotide 536 to the end of the coding region at nucleotide 839, amino acids 179 to 279 of the C1 protein) between the CaMV 35S promoter and the CaMV 35S terminator;

Please replace the paragraph on page 11 beginning at line 11 with the following rewritten paragraph:

Figure 28 shows a schematic diagram of the plasmid pOH007 as used in the present invention. The plasmid is a double construct in pBINplus containing the chimeric promoter (SEQ ID NO: 19), a GUS intron (Vancanneyt *et al.*, 1990) and a nos terminator, and also containing the translational fusion between the ohpR coding sequence (nucleotide 295 to nucleotide 1035 of SEQ ID NO: 19) and part of the C1 cDNA (from the PstI site at nucleotide 674 to the end of the coding region at nucleotide 839, amino acids 219 to 279 of the C1 protein) between the CaMV 35S promoter and the CaMV 35S terminator.

Please replace the paragraph on page 17 beginning at line 33 with the following rewritten paragraph:

Preferably, the inducer which causes modulation of expression of the nucleic acid sequence is a chemical compound, such as OHP, 2-hydroxy cinnamic acid,

toluene, ~~be~~zene benzene, n-hexadecane or a functional equivalent of either. The inducer may also, however, be a protein or nucleic acid sequence, depending on the complementary domain of the regulator sequence. The 5' regulatory regions of the second nucleotide sequence may suitably comprise one or more response elements, each being necessary for complementary binding of an appropriate domain or other portion of the regulator sequence.

Please replace the paragraph on page 20 beginning at line 32 with the following rewritten paragraph:

The plasmid pSK-59 (Figure 4) was digested with Xho1 and Sal1, the 414bp operator region was gel-purified and ligated with ~~p~~BS52040 pSK52040 (Figure 6 5) which had been digested with XhoI and phosphatased. The resulting plasmid was named pSK58040 (Figure 7 6).

Please replace the paragraph on page 21 beginning at line 4 with the following rewritten paragraph:

The three oligonucleotides CaMVop2 (SEQ ID NO: 14), CaMVop3 (SEQ ID NO: 15) and CaMVop4 (SEQ ID NO:16) were annealed in equimolar amounts (500 pmole each primer) and diluted tenfold. 5 μ l of this dilution were used as a template for a PCR reaction (50 μ l total) catalysed by a proof-reading Taq polymerase to generate double stranded product. The PCR product was resolved on an 8% polyacrylamide gel. The 125 bp PCR product was excised and purified using techniques described in Sambrook et al (1989). 1 μ l of the total eluted double stranded DNA solution (50 μ l) was used as a template in a PCR reaction (50 μ l total) primed by oligonucleotide primers CaMVopF1 (SEQ ID NO: 17) and CaMVopR1 (SEQ ID NO: 18) and catalysed by a proof-reading Taq polymerase. The PCR product from this reaction was digested to completion with EcoRV and BamHI and the 133 bp restriction fragment ligated with plasmid pDV35S1 (Figure 5 7) similarly digested to completion with EcoRV and .the resulting construct was named pDV60 (Figure 8). The inserted region was sequenced. Plasmid pDV60 (Figure 8) was digested with XhoI and BamHI. The 476bp synthetic promoter restriction fragment (SEQ ID NO: 19) was gel purified as described above and ligated into pSK52040 (Figure 6 5)

similarly digested with XhoI and BamHI. This plasmid was named pSK60040 (Figure 9). The chimeric promoter in SEQ ID NO: 19 contains a 36 bp region of the ohp operon (from nucleotide 1225 to nucleotide 1260) inserted into the CaMV 35S promoter at nucleotide 21.

Please replace the paragraph on page 22 beginning at line 4 with the following rewritten paragraph:

pDV35S1 (Figure 5 7) was digested with HindIII and SacI and the 668 bp fragment containing the CaMV 35S promoter/terminator was gel-purified and ligated with pUCAP (Figure 12) which was digested with HindIII and SacI. The resulting construct was named pDV35S2 (Figure 13).

Please replace the paragraph on page 22 beginning at line 22 with the following rewritten paragraph:

Plasmid pSK58040 (Figure 7 6) was digested to completion with HindIII and SmaI and the 2837 bp fragment containing the CaMV 35S promoter-GUS-nos terminator was gel-purified and ligated into pBINplus (Figure 17) similarly digested with HindIII and SmaI. The resulting plasmid was named pBNP58040 (Figure 18).

Please replace the paragraph on page 24 beginning at line 30 with the following rewritten paragraph:

Following transformation, the transformed cell or plant tissue is selected or screened by conventional techniques. The transformed cell or plant tissue contains the chimeric DNA sequences discussed above and is ~~the~~ then regenerated, if desired, by known procedures. The regenerated plants are screened for transformation by standard methods. Progeny of the regenerated plants is continuously screened and selected for the continued presence of the integrated DNA sequence in order to develop improved plant and seed lines. The DNA sequence can be moved into other genetic lines by a variety of techniques, including classical breeding, protoplast fusion, nuclear transfer and chromosome transfer.

Please replace the paragraph on page 25 beginning at line 9 with the following rewritten paragraph:

Transient expression assays of the gene expression cassette ~~was~~ were essentially performed as described by Kapila *et al.*, (1997), Rossi *et al.*, (1993), Twell *et al.*, (1989), Goff *et al.*, (1990), Roth *et al.*, (1991) and Tuerck *et al.*, (1994).